

Assessment of the presence of antibiotic residues in meat from small-scale broiler farms in the Pwani region of Tanzania

Rogia Saidath Adeline Gomez ^{a,*}, Isabella Mandl ^b, Said Hemed Mbaga ^a

^aDepartment of Animal, Aquaculture and Range Sciences, Sokoine University of Agriculture, Morogoro, Tanzania

^bDepartment of Botany and Biodiversity Research, University of Vienna, Austria

Abstract

The unregulated use of veterinary drugs, particularly antibiotics, on Tanzanian broiler farms is widespread. Still, little attention has been paid to the factors that promote the presence of antibiotic residues in broiler meat. This study aimed to identify rearing practices likely to favour the presence of antibiotic residues and to determine the presence of residues of commonly used antibiotics in broiler meat. The length of the rearing period, poultry pathologies and the type of veterinary medicines used were studied on 78 farms in the Pwani region (Tanzania). The results showed that infectious coryza, coccidiosis, chronic respiratory diseases, salmonellosis and omphalitis were the most common diseases. Of the veterinary medicines used to treat these diseases, antibiotics were the most widely used (69%), mainly enrofloxacin (ENO), limoxin (LX), oxytetracycline (OTC), tylosin (TS) and tyloxin (TDX). Eight farms were selected to take broiler meat samples for laboratory analysis of residues of these five antibiotics. These analyses revealed that all but one of the samples were negative. Samples taken from four-week-old birds tested positive for another antibiotic, sulfamethazine (SF), which was not initially one of the five antibiotics selected for this study. This study highlighted the significant use of veterinary drugs in response to various diseases. The absence of antibiotic residues can be attributed to the age of the animals sampled, as at over 5 weeks they would have had time to eliminate the drugs. A rearing period of four weeks is insufficient and would be a factor favouring the presence of drug residues in broiler meat.

Keywords: Antibiotics, avian diseases, LC-MS/MS, poultry meat, rearing length

1 Introduction

Animal health involves the use of either naturally produced or synthesised antibiotics to prevent (prophylaxis), treat (antibiotic therapy) or control diseases (metaphylaxis) (Taylor & Walker, 1956; Kantati, 2011; Oufella & Smail, 2012; Buket *et al.*, 2013; Mensah *et al.*, 2014; Mohamed Said, 2015; Okombe *et al.*, 2016; Prajapati *et al.*, 2018; Zamouma *et al.*, 2020; Moga *et al.*, 2021; Azabo *et al.*, 2022). They are also used as growth stimulants in poultry farming to increase productivity while reducing the stress associated with farming (zootechnical activities, vaccination) and the impact of disease (Zerbo, 2014; Agbodossindji *et al.*, 2018; Ahmed et Ben Hamida, 2019).

Although there are several families of antibiotics, the most commonly used in poultry farming are tetracyclines, B-

lactams, macrolides, quinolones and sulphonamides (Mes-sai, 2006; Chafer-Pericas *et al.*, 2010; Oufella & Smail, 2012; Renaud, 2012; Mohamed Said, 2015; Ramatla *et al.*, 2017; Ahmed & Ben Hamida, 2019; Kumar *et al.*, 2020; Moga *et al.*, 2021).

Antibiotics offer many advantages, but their inappropriate and uncontrolled use has harmful consequences (antibiotic resistance, presence of antibiotic residues in animal products, environmental contamination, etc.) (Hinton, 1988; Parent, 2009; Zerbo, 2014; Bagré *et al.*, 2015; FAO, 2015; Gaudin, 2016; Okombe *et al.*, 2016; Ramatla *et al.*, 2017; Agbodossindji *et al.*, 2018; Vana *et al.*, 2020; Mdegela *et al.*, 2021; Sangeda *et al.*, 2021).

Furthermore, uncontrolled use of these veterinary products has been observed in Tanzanian poultry farming, particularly commercial poultry (broilers and layers), despite the existence of regulations on the use of antibiotics. This

* Corresponding author: rogiaomez20@gmail.com

is primarily due to easy access to antibiotics, inappropriate use, incorrect dosage, and non-compliance with withdrawal periods, which is the minimum period between the administration of a drug to poultry and its slaughter (Mdegela *et al.*, 2021; Sangeda *et al.*, 2021). Antibiotic residues in poultry products have already been reported in several African countries (Egypt, Ethiopia, Ghana, Kenya, South Africa, Sudan, and Tanzania). This increased and uncontrolled use of antibiotics exposes consumers of poultry products to health risks (hypersensitivity reactions, cancer, bacterial resistance in humans, toxicity, allergies, changes in the intestinal flora, etc.) (Wouembe, 2013; Zerbo, 2014; Mohamed Said, 2015; Gaudin, 2016; Vana *et al.*, 2020). In addition, in developing countries such as Cameroon, Ghana and Kenya, the absence, inefficiency or lack of regulations governing the circulation and use of veterinary products contributes to the growing and uncontrolled use of antibiotics (Oufella & Smail, 2012; FAO, 2015; Ahmed & Ben Hamida, 2019; Kimera *et al.*, 2020; Vana *et al.*, 2020; Mdegela *et al.*, 2021; Mshana *et al.*, 2021; Sangeda *et al.*, 2021; Mouiche *et al.*, 2022; Mudenda *et al.*, 2022). To protect public health, sanitary control of animal products (microbiological contaminants, antibiotic residues, etc.) is essential. Similar considerations apply to the development and effective implementation of legislation and policies aimed at controlling the circulation, quality and use of antibiotics (Mensah *et al.*, 2014; Kimera *et al.*, 2020; Zamouma *et al.*, 2020; Oyedeji *et al.*, 2021).

Many studies, including those carried out in Tanzania, have focused on the detection of antibiotic residues in poultry meat and their possible contribution to the development of microbial resistance (Nonga *et al.*, 2009; Nonga *et al.*, 2010; Vana *et al.*, 2020; Mdegela *et al.*, 2021; Sangeda *et al.*, 2021; Ulomi *et al.*, 2022; Getahun *et al.*, 2023; Mohammed *et al.*, 2023; Nouri and Salehi, 2023; Hedges *et al.*, 2024; Marsini *et al.*, 2024; Tek *et al.*, 2024). However, little has been reported on the contribution of farming practices (inappropriate use, incorrect dosage, non-compliance with withdrawal periods) to the use of antibiotics and the presence of antibiotic residues in poultry meat. In addition, previous studies (Gomez & Mbaga, 2023; Gomez & Mbaga, 2024) have shown an average level of biosecurity and a high cost of treatment for avian diseases on broiler farms in the Pwani region, suggesting that these farmers purchase and use large quantities of veterinary medicines. The aim of this study was, therefore, to determine the presence of residues of the five antibiotics commonly used on these farms in broiler meat and to identify the farming practices likely to lead to contamination of broiler meat by antibiotic residues in the study region.

2 Materials and methods

2.1 Study area

This study was carried out in the Pwani region, specifically in Kibaha Council Town and the adjacent town of Mlandizi. In Tanzania, commercial poultry farming is mainly practised in Dar es Salaam, Pwani, Arusha, Mwanza and Tanga regions, with the Pwani region having the highest number of commercial poultry farms (layers or broilers) (FAO, 2007).

Geographically, situated in the middle Eastern side of Tanzania's Mainland, the Coast or Pwani Region is between S6°-8° and E37°30-40. Dar es Salaam Region and the Indian Ocean are boarded by Pwani Region in the East, North by Tanga Region, South by Lindi and West by Morogoro Region. It is in these areas that small and medium-sized broiler rearing is most widespread in the Pwani region.

2.2 Choice of sampling method

To identify the farms where broiler meat samples were acquired in the Pwani region, the non-probability sampling method known as "snowball sampling" (Orounladji *et al.*, 2022) was used. This sampling method was chosen due to the lack of precise statistical data on the broiler population and the number of broiler farmers in Tanzania (FAO, 2007; Msoffe *et al.*, 2018; EKN, 2020). It is a sampling method based on networking (Kone *et al.*, 2018). As a result, the first farmer to be surveyed then nominated other farmers in the same area who could also be surveyed (Marpsata & Razafindratsima, 2010; Kouassi *et al.*, 2019).

2.3 Sampling design

This study is subdivided into three parts: (i) the field survey (which collected information on the respondents' rearing practices); (ii) the collection of broiler meat samples (for laboratory analysis); and (iii) the laboratory analysis phase (for the presence of residues of the top five antibiotics most commonly used by respondents in the study area).

2.4 Phase one: Field survey

A total of 78 broiler farmers were interviewed individually using a questionnaire. The questionnaire administered included questions relating to the length of time broilers have been reared in the Pwani region, the diseases affecting broiler flocks and the veterinary medicines used to treat the diseases. Additional questions were also administered to farmers in the Pwani region, such as the type of treatment (no treatment of diseases, treatment by a veterinarian, endogenous treatment and self-medication) practised by respondents, the

reasons for using veterinary medicines, and the frequency of use of veterinary medicines. The survey was conducted over three months (November 2021 to January 2022).

2.5 Phase two: Collection of broiler meat samples

Based on the preliminary results of our questionnaires, we selected eight farms for meat sampling: the farms selected for broiler meat collection were those that used at least two of the five antibiotics considered in the search for antibiotic residues in meat and that had birds ready for disposal. On the eight farms selected according to the above two characteristics, a sample of three broilers aged between four and six weeks per farm was taken, giving a sample size of $n = 24$. After the broilers had been slaughtered, 100 g of the breast from each chicken was taken, stored in a zip-lock bag, labelled accordingly and then immediately stored in a cooler with ice cubes before being transported to the laboratory (Nonga et al., 2009).

2.6 Phase three: Laboratory analysis

2.6.1 Laboratory analysis method

The samples were taken to the Government Chemist Laboratory Authority-Tanzania (GCLA-Tanzania) for analysis. The meat samples collected were analysed using liquid chromatography-tandem mass spectrometry (LC-MS/MS). In this study, and given the circumstances in the field, this method was only used to screen for the antibiotics under investigation (ENO, OTC, LX, TS and TDX). LC-MS/MS was chosen because it is a modern, rapid method that is widely used for screening for tetracyclines, quinolones and macrolides (the families to which the antibiotics investigated in this study belong) and for detecting multi-residues (Granelli et al., 2009; Parent, 2009; Chafer-Pericas et al., 2010; Kantati, 2011; Renaud, 2012; Zamouma et al., 2020).

2.6.2 Reagents and devices used in laboratory analysis

The reagents used for antibiotic residue detection in these broiler breast meat samples are 70 % methanol (HPLC grade), acetonitrile (ACN) (HPLC grade), formic acid, Milli Q water and 0.1 M EDTA. Apparatus and supplies used during this laboratory analysis are: weigh balance; Vortex machine; nitrogen evaporator and nitrogen generator; centrifuge machine 4000 rpm and microcentrifuge 13000 rpm; 1000 μ l micropipette and 1000 μ l sterile tips; laboratory blender; scissor; tongs; timer; conical centrifuge tubes 15 ml and 50 ml, beaker 50 ml, 100 ml and 500 ml, Eppendorf tubes 1500 μ l, 1500 μ l LC vial, test tubes, graduated measuring cylinder 10 ml, 50 ml and 100 ml.

2.6.3 Sample preparation

The samples were extracted at one go with 70 % methanol, diluted with distilled water and injected into the LC-MS/MS following the procedure of Granelli et al. (2009) and adapted for this study. The chromatography was performed on a Hypersil Gold aQ C18 column (2.1*100 mm, 1.9 μ m). For the mobile phase, 0.1 % formic acid in Milli Q water (mobile phase A) and formic acid in acetonitrile (mobile phase B) were used. The gradient was from 5 % B to 100 % B in 13 min, was maintained at 100 % B until 20 min and returned to 5 % B at 30 min. Each injection lasted 7 minutes and the flow rate and injection volume were 0.3 ml min⁻¹ and 10 μ l, respectively. The temperature of the column was 40 °C. To obtain the 0.1 M EDTA, 37.224 g EDTA was dissolved in water in a 1000 ml volumetric flask. Then, the flask was filled with distilled water until the graduation limit was reached. The 70 % methanol was obtained by mixing 350 ml Methanol with 150 ml water in a 500 mL volumetric flask. The 0.1 % Formic acid used in mobile phase A was obtained by adding 500 μ l of formic acid to 499.5 ml of distilled water. 0.1 % formic acid used in mobile phase B was obtained by adding 500 μ l of formic acid to 499.5 ml of acetonitrile. In summary, 200 μ l of 0.1 M EDTA (ethylenediaminetetraacetic acid) was added to 2 g of homogenised tissue. After spiking, the antibiotics were extracted from the tissue using 5 ml of 70 % methanol. After extraction, the samples were centrifuged at 4,000 rpm for 10 minutes. The supernatant was concentrated to 1 ml in the nitrogen evaporator; then transferred to a 1500 ml Eppendorf tube and centrifuged again at 13,000 rpm for 5 minutes. After centrifugation, 800 ml of the contents were transferred to a 1,500 ml Eppendorf tube and vortexed. Finally, 10 ml of the contents of the 1500 ml vial were removed and injected into the LC-MS/MS machine for analysis. In addition to the LC vials containing the extracts of the 24 tissue samples to be analysed, other LC vials containing a spiked sample and a blank sample were added. The spiked sample was prepared following the same procedure but was free of animal tissue. A blank sample was only solvents without adding a sample was used as method blank. 200 μ l of 0.1 M EDTA was taken to a 15 ml conical centrifuge tube and put in the dark for 15 mins then followed all other steps as a sample.

2.7 Statistical analysis

The analysis of the data in this study focused on descriptive statistics, as these correspond best to the results of the survey and laboratory analysis. The data collected was stored and processed in an Excel spreadsheet (Microsoft Corporation, WordNet 3.0 Copyright 2006 by Princeton University, Version 2023).

3 Results

Overall, chicken rearing (the main source of income on the farms surveyed) was combined with other income-generating activities such as farming. On average, farmers in the region were 52 years old and had been rearing broilers for about 9 years. The domestic animals reared on the surveyed farms were not limited to broilers. Local chickens, laying hens and other animals such as cattle, sheep and rabbits were also present.

Table 1 indicates the avian diseases identified in this study. The most common were infectious coryza, coccidiosis, chronic respiratory disease, salmonellosis, and omphalitis (Table 2). Most of these diseases occurred during the first two weeks of rearing, except for coccidiosis and Newcastle disease, which could happen at any time during the rearing period. According to the interviewees, the type of treatment frequently used was treatment prescribed by a veterinary officer or self-medication (use of a veterinary product on the breeder’s own initiative to treat diseases without consulting an animal health officer).

Table 1: Categorisation of diseases found on the 78 surveyed broiler farms in the Pwani region (Tanzania).

Cause of disease	Disease
Bacteria	Infectious coryza
	Chronic respiratory disease
	Salmonellosis
Virus	Infectious bursal disease or Gumboro
	Newcastle Disease
Parasites	Coccidiosis
Mismanagement	Omphalitis
Nutritional deficiencies	Deficiency of minerals

Table 2: Categorisation of diseases found on the 78 broiler farms surveyed in the Pwani region (Tanzania).

Disease	Freq.*	Treatment†
Infectious coryza	54	1, 3
Coccidiosis	43	1, 3
Chronic respiratory disease	15	1, 2
Salmonellosis	14	1, 3
Omphalitis	10	1, 3
Deficiency of minerals	4	1, 3
Infectious bursal disease or Gumboro	3	1, 2, 3
Newcastle Disease	1	2, 3

*Frequency of citations; †0: no treatment; 1: veterinarian; 2: traditional treatment; 3: self-medication.

The treatment of these diseases involved the use of veterinary medicines, especially antibiotics (69 % of veterinary drugs used) by most of the respondents (Fig. 1A and 1B). These veterinary drugs were used for prophylaxis (19 % of veterinary drug use) or antibiotic therapy (81 %).

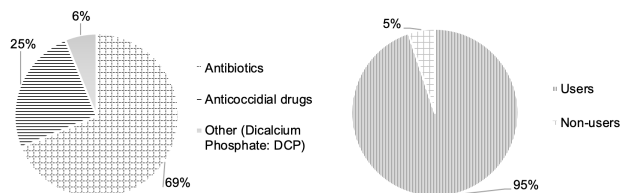


Fig. 1: Type of veterinary drugs used (left) & rate of veterinary medicine users (right) on the 78 broiler farms surveyed in the Pwani region (Tanzania).

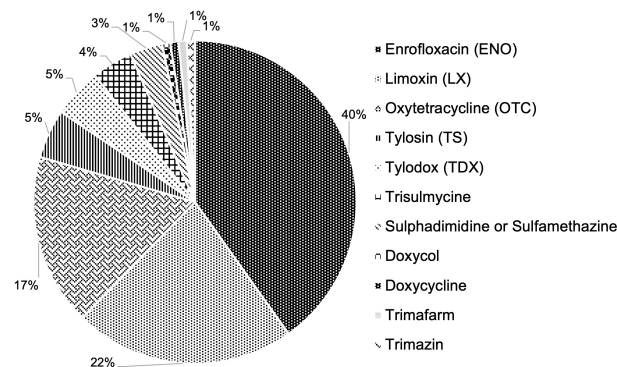


Fig. 2: Types of antibiotics used on the 78 broiler farms surveyed in the Pwani region (Tanzania).

The antibiotics commonly used by respondents were enrofloxacin (ENO), limoxin (LX), oxytetracycline (OTC), tylosin (TS) and tylodox (TDX) (Fig.2). As far as anticoccidials were concerned, the one most frequently used was amprolium (Fig.3).

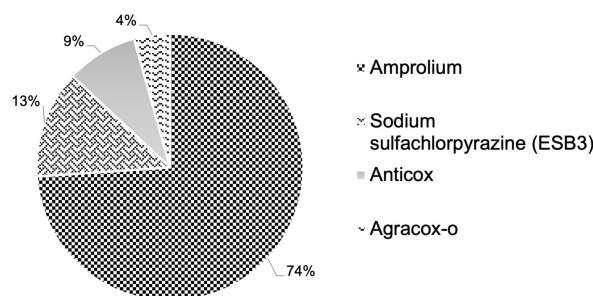


Fig. 3: Anticoccidial used on the 78 broiler farms surveyed in the Pwani region (Tanzania).

The diseases treated by the five commonly used antibiotics (ENO, LX, OTC, TS and TDX) are shown in Table 3. It should be noted that these antibiotics were used to treat bacterial (infectious coryza, etc.), parasitic (coccidiosis) and viral (Gumboro) diseases.

Table 3: Frequency of use of the top five used antibiotics by type of disease on the 78 broiler farms surveyed in the Pwani region (Tanzania).

Antibiotics	Disease*	Freq. [†]
Enrofloxacin (ENO)	Infectious coryza (34)	3 to 1
	Chronic respiratory disease (8)	3
	Used during brooding (4)	3
	Gumboro* (1)	3
Limoxin (LX)	Infectious coryza (20)	3 to 1
	Chronic respiratory disease (4)	3
	Coccidiosis [‡] (1)	1
	Gumboro [‡] (1)	3
	Omphalitis (Navel disease) (1)	2
Oxytetracycline (OTC)	Used during brooding (12)	3
	Infectious coryza (3)	3
	Omphalitis (Navel disease) (1)	2 to 1
Tylosin (TS)	Infectious coryza (3)	3
	Coccidiosis [‡] (2)	3
	Salmonellosis (1)	3
	Used during brooding (1)	3
Tyloxox (TDX)	Infectious coryza (3)	1 to 2
	Chronic respiratory disease (2)	3
	Used during brooding (1)	3

*Number of responses in brackets; [†]Frequency of use of antibiotics 1: very frequent; 2: moderate; 3: occasionally; [‡]Non-bacterial diseases not effectively treated by antibiotics.

For laboratory tests to detect residues of the five antibiotics (ENO, LX, OTC, TS and TDX), meat samples were taken from broilers aged four weeks (n = 3) on one selected farm and five to six weeks (n = 21) on the remaining seven farms. These analyses showed that all 24 samples from the eight farms studied were negative for the five antibiotics in searching. However, samples from four-week-old birds tested positive for another antibiotic, sulfamethazine (SF), which was not initially one of the five antibiotics selected for this study.

4 Discussion

In this study, the field survey revealed that broiler farmers in the Pwani region were mainly confronted with respiratory diseases (infectious coryza and chronic respiratory disease), followed by enteric diseases (coccidiosis), systemic diseases

(salmonellosis) and Omphalitis. Our results are similar to those of MLD (2006), Nonga *et al.* (2009), Nonga *et al.* (2010), Pare (2012), Swai *et al.* (2013), Msoffe *et al.* (2018), Chota *et al.* (2021), Mdegela *et al.* (2021), et Mouiche *et al.* (2022), who noted that the diseases cited in this study were among the most common poultry diseases. Viral diseases (Newcastle disease and Gumboro) represented only a small proportion of the diseases mentioned by respondents, mainly due to compliance with the vaccination plan for broilers against these diseases (Gomez and Mbaga, 2023). This considerable susceptibility of broilers could be explained by several factors, such as the quality of day-old chicks (DOC), rearing conditions and genetics. Omphalitis (inflammation of the navel) only occurred in the event of poor hygiene conditions in hatcheries, transport and chick starting (ASA, 2005; Kariuki *et al.*, 2023). According to Rahmatallah *et al.* (2018), poor DOC quality leads to infection of the yolk sac and a high prevalence of respiratory diseases. According to Gomez & Mbaga (2023), chicken farmers in the Pwani region faced a high mortality rate (11.7%) with average biosecurity practice within the surveyed farms, suggesting a high prevalence of avian diseases. It should also be noted that bred broilers are exotic breeds often poorly adapted to African environmental conditions (Pare, 2012).

The high prevalence of avian diseases in this study area could be the main reason for the increased use of veterinary medicines. It seems that the majority of respondents used veterinary medicines during the rearing period. Use was either prophylactic (especially at the start of rearing) or therapeutic. This result is consistent with the results of Nonga *et al.* (2010), Rahmatallah *et al.* (2018), Kimera *et al.* (2020), Vana *et al.* (2020), Chota *et al.* (2021), Mdegela *et al.* (2021) et Ulomi *et al.* (2022), where many livestock keepers used veterinary medicines for prophylaxis, metaphylaxis and antibiotic therapy in several African countries. According to Kariuki *et al.* (2023), some DOC vendors tend to encourage poultry keepers to use veterinary drugs, rather than emphasizing adherence to biosecurity measures to prevent omphalitis and early mortalities. Another factor that could influence the use of veterinary medicines in this study is the easy accessibility and uncontrolled use of these products on the market, which would favour self-medication, which was the most common type of treatment used by the respondents. Self-medication (use of a veterinary product on one's own initiative to treat diseases without consulting an animal health worker) could be conditioned by easy access (without prescription or restriction) to veterinary drugs, knowledge of the brands of veterinary drugs, the lack of regulations on the use of veterinary drugs in several African countries, the lack of extension and veterinary advisory ser-

vices, and the dependence of livestock farmers on their personal experience (Nonga *et al.*, 2009; Nonga *et al.*, 2010; Okombe *et al.*, 2016; Mouiche *et al.*, 2022; Mudenda *et al.*, 2022; Vana *et al.*, 2020; Ulomi *et al.*, 2022; Kariuki *et al.*, 2023). This situation (self-medication) could partly explain the use of different veterinary medicines for the same diseases. For example, the results showed that coccidiosis was treated either with antibiotics (ENO, LX, TS) or with antiprotozoal (amprolium, ESB3, anticox). While there was a significant prevalence of diseases on the farms surveyed associated with the remarkable use of veterinary medicines, all of the respondents said they occasionally used veterinary medicines. This could mean that none of the interviewees recorded the care of the birds and were unaware of the frequency of use of these veterinary drugs. According to Mouiche *et al.* (2022), the absence or poor management of treatment records would lead farmers to misuse veterinary medicines.

Laboratory results exhibited that all samples were negative for the five antibiotics concerned (ENO, LX, OTC, TS and TDX). However, samples from four-week-old birds tested positive for another antibiotic, sulfamethazine (SF), which was not initially one of the five antibiotics selected for this study. According to the literature (Fei *et al.*, 2023; Chandrakar *et al.*, 2023; Kamouh *et al.*, 2023; Sani *et al.*, 2023; Abdel-Wahhab *et al.*, 2024), the five antibiotics researched and the one tested positive in broiler meat in this study are among the antibiotics from the families (tetracyclines, quinolones, beta-lactams, sulphonamides, aminoglycosides and macrolides) most commonly used in poultry production and whose residues are the most sought-after in animal products.

What's more, the laboratory where the samples were analysed had developed a special adaptation for the LC-MS/MS method which, after injecting the samples into the LC-MS/MS machine, made it possible to read all the chemical compounds contained in each sample. This made it possible to detect the presence of another antibiotic in samples from four-week-old broilers. Remember that sulphonamides, used in the treatment and prevention of diseases such as infectious coryza, avian typhoid and coccidiosis, have a low purchase cost, which makes them easily accessible to farmers and could lead to their inappropriate use. Uncontrolled use of sulphonamides could increase the incidence of bacteria that are resistant to them and the presence of their residues in meat if their withdrawal periods are not respected (Helal *et al.*, 2024; Irani *et al.*, 2024).

The negative results of the samples analysed in this study could be explained by the absence of use of these antibiotics by the farmers during the periods preceding the purchase and slaughter of these broilers on the one hand, and

by the age at which these animals were slaughtered on the other. The duration of broiler rearing observed in this study was four weeks, which is short compared with the standard six weeks (39–42 days) required for broiler rearing (Guérin, 2004). Furthermore, most of these antibiotics are prescribed for a treatment period of three to five days, and their withdrawal times vary from one antibiotic to another. For example, the withdrawal times for ENO, OTC and TS are seven and three days respectively after the last dose administered (Khatun *et al.*, 2018; Chota *et al.*, 2021). In contrast, sulphonamides, the antibiotic family to which FS belongs, have a withdrawal time of up to 16 days after the last administration of the antibiotic (Chota *et al.*, 2021), which could justify its presence in the meat of animals aged four weeks at the time of laboratory analysis.

These results reveal, on the one hand, that the four-week rearing period practised by farmers is insufficient to raise broilers properly and, on the other hand, that it is important to respect the withdrawal period for the various veterinary medicines. The farmers who kept broilers in stock for more than four weeks may have unknowingly allowed the withdrawal period, likely contributing to the non-detection of antibiotics in the laboratory samples. Hence the failure to detect antibiotics in laboratory samples. According to several authors (Nonga *et al.*, 2009; Nonga *et al.*, 2010; Zerbo, 2014; Ahmed & Ben Hamida, 2019; Mudenda *et al.*, 2022), failure to observe the withdrawal period for veterinary medicines exposes consumers of poultry products to health risks (allergies, poisoning, etc.). Non-compliance with the withdrawal period could be linked to a lack of knowledge or ignorance of its impact (contribution to antibiotic resistance, presence of ARs in meat and eggs, etc.). According to Prajapati *et al.* (2018), the short lifespan of broilers could also encourage farmers not to comply with waiting times for veterinary medicines. The other reason is that farmers are forced to dispose of (early sell) broilers when they are less than five weeks old, due to rising production costs (steadily increasing costs of feed and rearing inputs), while the selling price of broilers has remained stagnant over the years. Ultimately, this could hurt the general health of the population (hypersensitivity reactions, cancer, bacterial resistance in humans, toxicity, allergies, changes in the intestinal flora, etc.) (Mohamed Said, 2015; Gaudin, 2016; Vana *et al.*, 2020), requiring holistic intervention.

5 Conclusion and recommendation

This study exhibited the extensive use of veterinary medicines, especially antibiotics, to combat the avian diseases that were rife in the study area. Several factors predispose to

such situations, such as the accessibility of veterinary medicines, self-medication and the ineffectiveness of regulations on the use of veterinary medicines. The four-week rearing period seems inadequate, leading to non-compliance with waiting periods for the various veterinary drugs, and is a factor that favours the presence of drug residues in broiler meat. Consequently, rearing broilers for more than four weeks, assuming that prophylactic measures and antibiotic treatments have been taken earlier (from zero to four weeks), presents little danger, as it will allow the antibiotics to be withdrawn and the meat product to be made fit for consumption. It is recommended that broiler farmers be made aware of the application of biosecurity, the use of veterinary medicines and their impact in the event of misuse.

Conflict of interest

The authors declare that they have no conflict of interest.

Acknowledgements

The authors thank the Partnership for Skills in Applied Sciences, Engineering & Technology Regional Scholarship and Innovation Fund, the Department of Animal, Aquaculture and Range Sciences, the Mawazo Institute, the Southern African Centre for Infectious Disease Surveillance Foundation for One Health, as well as the Livestock Extensions and all the respondents of study areas and Dr Boko Michel Orounladji.

References

- Abdel-Wahhab, R. M., Hussein, H. K., ElBarbary, M., & Mahmoud, H. A. (2024). Overview On Some Antibiotic Residues in Poultry Meat. *Alexandria Journal of Veterinary Sciences (AJVS)*, 81(1), 157–162. DOI:10.5455/ajvs.196539.
- Agbodossindji, A. S., Mensah, S. E. P., Adjahoutonon, K. Y. K. B., Attakpa, E., Koudandé, O. D., & Mensah, G. A. (2018). Evaluation of antibiotic residues in imported frozen chicken thigh muscles marketed in Southern Benin. *African Union Inter-African Bureau for Animal Resources. Bulletin of Animal Health and Production in Africa*, 66(4), 751–758.
- Ahmed, F. Z., & Ben Hamida, H. (2019). *Détection des résidus d'antibiotiques dans la viande du poulet de chair dans la région de M'sila*. Mémoire présenté pour l'obtention Du diplôme de Master Académique. Université Mohamed Boudiaf - M'sila, Faculté des Sciences. Pp. 68.
- ASA. (American Soybean Association) (2005). *Handbook on Poultry Diseases*. 2nd Edition. 210p.
- Azabo, R., Dulle, F., Mshana, S. E., Matee, M., & Kimera, S. (2022). Antimicrobial use in cattle and poultry production on occurrence of multidrug resistant *Escherichia coli*. A systematic review with focus on sub-Saharan Africa. *Frontiers in Veterinary Science*, 9:1000457. doi: 10.3389/fvets.2022.1000457.
- Bagré, T. S., Samandoulougou, S., Traoré, M., Illy, D., Bsadjo-Tchamba, G., Bawa-Ibrahim, H., Bouda, S. C., Traoré, A. S., & Barro, N. (2015). Détection biologique des résidus d'antibiotiques dans le lait et produits laitiers de vache consommés à Ouagadougou, Burkina Faso. *Journal of Applied Biosciences*, 87, 8105–8112. <http://dx.doi.org/10.4314/jab.v87i1.11>.
- Buket, E., Onurdağ, F. K., Demirhan, B., Özgacar, S. Ö., Öktem, A. B., & Abbasoğlu, U. (2013). Screening of quinolone antibiotic residues in chicken meat and beef sold in the markets of Ankara, Turkey. *Poultry Science Association Inc.*, 92, 2212–2215. <http://dx.doi.org/10.3382/ps.2013-03072>.
- Chafer-Pericas, C., Maquieira, A., & Puchades, R. (2010). Fast screening methods to detect antibiotic residues in food samples. *Trends in Analytical Chemistry*, 29(9), 1038–1049. doi:10.1016/j.trac.2010.06.004.
- Chandrakar, C., Shakya, S., Patyal, A., Bhonsle, D., & Pandey, A. K. (2023). Detection of antibiotic residues in chicken meat from different agro-climatic zones of Chhattisgarh, India by HPLC-PDA and human exposure assessment and risk characterization. *Food Control*, 148. <https://doi.org/10.1016/j.foodcont.2023.109667>.
- Chota, A., Kitojo, O., & Ngongolo, K. (2021). Knowledge on diseases, practices, and threats of drugs residues in chicken food chains in selected districts of Dodoma region, Tanzania. *Journal of Applied Poultry Research*, 30:100186. <http://creativecommons.org/licenses/by-nc-nd/4.0/>.
- EKN. (2020). Market Trends and Consumer Behaviours and Preferences in The Tanzania Poultry Subsector: An Analytical Report with Recommendations for the Public and Private Sectors. Funded by the Embassy of the Kingdom of the Netherlands Dar es Salaam, Tanzania and conducted jointly by Ringo, E. J., & Lekule, F. P. from Match Maker Associates Limited. 50.
- FAO. (2007). Poultry sector country review. The structure, marketing, and importance of the commercial and village poultry industry: an analysis of the poultry sector in Tanzania. 61.
- FAO. (2015). The white meat value chain in Tanzania. A report from the Southern Highlands Food Systems Programme. pp.138.

- Fei, Z., Song, S., Gao, J., Song, Y., Xiao, X., Yang, X., Jiang, D., & Yang, D. (2023). Antibiotic residues in chicken meat in China: Occurrence and cumulative health risk assessment. *Journal of Food Composition and Analysis*, 116. <https://doi.org/10.1016/j.jfca.2022.105082>.
- Gaudin, V. (2016). *Caractérisation de la performance et validation des méthodes de dépistage des résidus d'antibiotiques dans les denrées alimentaires*. Thèse présentée pour le grade de Docteur de l'Université de Rennes 1, Mention : Biologie, Sciences agricoles. Université Rennes 1. <https://tel.archives-ouvertes.fr/tel-01439202>.
- Getahun, M., Abebe, R. B., Sendekie, A. K., Woldeyohanis, A. E., & Kasahun A. E. (2023). Evaluation of Antibiotics Residues in Milk and Meat Using Different Analytical Methods. *International Journal of Analytical Chemistry*, 2023(1), 1–13. <https://doi.org/10.1155/2023/4380261>.
- Gomez, R. S. A., & Mbagi, S. H. (2023). Assessment of biosecurity practices adoption in broiler farms in east Africa: case study of pwani region in Tanzania. *Journal of Animal Health and Production*, 11(2), 155–164. <http://dx.doi.org/10.17582/journal.jahp/2023/11.2.155.164>.
- Gomez, R. S. A., & Mbagi, S. H. (2024). Classification and productivity of smallholder broiler farms in the Pwani region, Tanzania. *Journal of Animal Health and Production*, 12(1), 31–39. <http://dx.doi.org/10.17582/journal.jahp/2024/12.1.31.39>.
- Granelli, K., Elgerud, C., Lundström, A., Ohlsson, A., & Sjöberg, P. (2009). Rapid multi-residue analysis of antibiotics in muscle by liquid chromatography-tandem mass spectrometry. *Analytica Chimica Acta*, 637, 87–91. doi: 10.1016/j.aca.2008.08.025}.lititem Hedges, S., Pelligand, L., Chen, L., Seow, K., Hoang, T. T., Luu, H. Q., Dang, S. T. T., Pham, N. T., Pham, H. T. T., Cheah, Y. C., Wang, Y., Hurtaud-Pessel, D., Conan, A., Fournié, G., Blake, D., Tomley, F., & Conway, P. L. (2024). Antimicrobial residues in meat from chickens in Northeast Vietnam: analytical validation and pilot study for sampling optimisation. *Journal of Consumer Protection and Food Safety*, 1–10. <https://doi.org/10.1007/s00003-024-01478-9>.
- Helal, D., Abobakr, E. M., Arab, W. S., & Elrais, A. M. (2024). Safety evaluation of some retailed chicken meat products with special reference to their antibiotic residue in Egypt. *Benha Journal of Applied Sciences (BJAS)*, 9(3), 127–132.
- Hinton, M. H. (1988). Antibiotics, poultry production and public health. *World's Poultry Science Journal*, 44(1), 67–69. DOI: <https://doi.org/10.1079/WPS19880009>.
- Irani, A. A., Moghaddam, A. D., Hadi, S., & Hadi, V. (2024). Evaluation of some sulfonamide antibiotics residues in chicken meat samples using in-syringe counter current homogenous liquid-liquid extraction and magnetic deep eutectic solvent-based dispersive liquid-liquid microextraction and HPLC–DAD. *Microchemical Journal*, 201. <https://doi.org/10.1016/j.microc.2024.110518>.
- Kamouh, H. M., Abdallah, R., Kirrella, G. A., Mostafa, N. Y., & Shafik, S. (2023). Assessment of antibiotic residues in chicken meat. *Open Veterinary Journal*, 14(1), 438–448. DOI:10.5455/OVJ.2024.v14.i1.40.
- Kantati, Y. T. (2011). *Détection des résidus d'antibiotiques dans les viandes de bovins prélevées aux abattoirs de Dakar*. Memoire de Master Qualite des aliments de l'homme, Spécialité : Produits d'origine animale. Présenté et soutenu à l'Ecole Inter-Etats des Sciences et Médecine vétérinaires (EISMV) de Dakar, Sénégal. Pp. 49
- Kariuki, J. W., Jacobs, J., Ngogang, M. P., & Howland, O. (2023). Antibiotic use by poultry farmers in Kiambu County, Kenya: exploring practices and drivers of potential overuse. *Antimicrobial Resistance and Infection Control*, 12(3), 11p. <https://doi.org/10.1186/s13756-022-01202-y>.
- Khatun, R., Howlader, A. J., Ahmed, S., Islam, N., Alam, K., Haider, S., Mahmud, M. S., & Hasan, Md. A. (2018). Validation of the Declared Withdrawal Periods of Antibiotics. *Universal Journal of Public Health*, 6(1), 14–22. DOI:10.13189/ujph.2018.060103.
- Kimera, Z. I., Mshana, S. E., Rweyemamu, M. M., Mboera, L. E. G., & Matee, M. I. N. (2020). Antimicrobial use and resistance in food-producing animals and the environment: an African perspective. *Antimicrobial Resistance and Infection Control*, 9(37), 1–12. <https://doi.org/10.1186/s13756-020-0697-x>.
- Kone, G. A., Kouassi, G. F., Kouakou, N. D. V., & Kouba, M. (2018). Diagnostic of Guinea fowl (*Numida meleagris*) farming in Ivory Coast. *Poultry Science*, 97, 4272–4278.
- Kouassi, G. F., Koné, G. A., Good, M., & Kouba, M. (2019). Factors impacting Guinea fowl (*Numida meleagris*) production in Ivory Coast. *Journal of Applied Poultry Research*, 28, 1382–1388.
- Kumar, H., Bhardwaj, K., Kaur, T., Nepovimova, E., Kuc̣a, K., Kumar, V., Bhatia, S. K., Dhanjal, D. S., Chopra, C., Singh, R., Guleria, S., Bhalla, T. C., Verma, R., & Kumar, B. (2020). Detection of Bacterial Pathogens and Antibiotic Residues in Chicken Meat: A Review. *Foods*, 9, 1504. 35p. DOI:10.3390/foods9101504.

- Marsata, M., & Razafindratsima, N. (2010). Les méthodes d'enquêtes auprès des populations difficiles à joindre : introduction au numéro spécial. *Methodological Innovations Online*, 5(2), 3–16. doi:10.4256/mio.2010.0014.
- Marsini, M., Pawestri, W., Wati, A. K., Yanti, Y., & Utami, L. (2024). Detection of antibiotic residues in broiler chicken meat in traditional markets in Surakarta City. In: IOP Conference Series: Earth and Environmental Science: 2nd International Conference on Animal Research for Eco-Friendly Livestock Industry, 1292(1), 1–7. doi: 10.1088/1755-1315/1292/1/012021.
- Mdegela, R. H., Mwakapeje, E. R., Rubegwa, B., Gebeyehu, D. T., Niyigena, S., Msambichaka, V., Nonga, H. E., Antoine-Moussiaux, N., & Fasina, F. O. (2021). Antimicrobial Use, Residues, Resistance and Governance in the Food and Agriculture Sectors, Tanzania. *Antibiotics*, 10(454), 23p. <https://doi.org/10.3390/antibiotics10040454>.
- Mensah, S. E. P., Aboh, A. B., Salifou, S., Mensah, G. A., Sanders, P., Abiola, F. A., & Koudandé, O. D. (2014). Risques dus aux résidus d'antibiotiques détectés dans le lait de vache produit dans le Centre Bénin. *Journal of Applied Biosciences*, 80, 7102–7112.
- Messaï, A. (2006). *Analyse critique des pratiques de l'antibiothérapie en élevages avicoles*. Mémoire présenté pour l'obtention du diplôme de Magister en médecine vétérinaire, Option : pathologie, Spécialité : Aviculture et pathologie aviaire. Université Mentouri de Constantine - Faculté des Sciences. p 133.
- MLD. (2006). National Livestock Policy. Ministry of Livestock Development, Tanzania. p. 55.
- Moga, A., Vergara-Barberán, M., Lerma-García, M. J., Carrasco-Correa, E. J., Herrero-Martínez, J. M., & Simó-Alfonso, E. F. (2021). Determination of antibiotics in meat samples using analytical methodologies: A review. *Comprehensive Reviews in Food Science and Food Safety*, 20, 1681–1716. <https://doi.org/10.1111/1541-4337.12702>.
- Mohamed Said, R. (2015). *Etudes qualitatives et quantitatives des résidus d'antibiotiques dans la viande de volaille et les oeufs dans la région de la Mitidja et utilisation des probiotiques comme alternative*. Thèse de doctorat en Science, spécialité : Sciences Biologiques, soutenue à l'Université Mouloud Mammei de Tizi Ouzou, Algérie. 159p
- Mohammed, M. A., Khalafalla, F. A., & Abdel-Atty, N. S. (2023). Antibiotics in Poultry Meat and Products' Residual Levels and Implications to Public Health. *Journal of Veterinary Medical Research*, 30(1), 36–40. <https://doi.org/10.21608/jvrmr.2022.178509.1076>.
- Mouiche, M. M. M., Wouembe, F. D. K., Mpouam, S. E., Moffo, F., Djuntu, M., Toukam, C. M. W., Kameni, J. M. F., Okah-Nnane, N. H., & Awah-Ndukum, J. (2022). Cross-Sectional Survey of Prophylactic and Metaphylactic Antimicrobial Use in Layer Poultry Farming in Cameroon: A Quantitative Pilot Study. *Frontiers in Veterinary Science*, 9:646484, 11p. doi:10.3389/fvets.2022.646484.
- Mshana, S. E., Sindato, C., Matee, M. I., & Mboera, L. E. G. (2021). Antimicrobial Use and Resistance in Agriculture and Food Production Systems in Africa: A Systematic Review. *Antibiotics*, 10(8), 976. <https://doi.org/10.3390/antibiotics10080976>.
- Msoffe, G., Chengula, A., Kipanyula, M. J., Mlozi, M. R. S., & Sanga, C. A. (2018). Poultry Farmers' Information needs and Extension advices in Kilosa, Tanzania: Evidence from Mobile-based Extension, Advisory and Learning System (MEALS). *Library Philosophy and Practice (e-journal)*, 18. <https://digitalcommons.unl.edu/libphilprac/1710>.
- Mudenda, S., Malama, S., Munyeme, M., Hang'ombe, B. M., Mainda, G., Kapon, O., Mukosha, M., Yamba, K., Bumbangi, F. N., Mfuné, R. L., Daka, V., Mwenya, D., Mpundu, P., Siluchali, G., & Muma, J. B. (2022). Awareness of Antimicrobial Resistance and Associated Factors among Layer Poultry Farmers in Zambia: Implications for Surveillance and Antimicrobial Stewardship Programs. *Antibiotics*, 11(3), 383. <https://doi.org/10.3390/antibiotics11030383>.
- Nonga, H. E., Mariki, M., Karimuribo, E. D., & Mdegela, R. H. (2009). Assessment of Antimicrobial Usage and Antimicrobial Residues in Broiler Chickens in Morogoro Municipality, Tanzania. *Pakistan Journal of Nutrition*, 8(3), 203–207.
- Nonga, H. E., Simon, C., Karimuribo, E. D., & Mdegela, R. H. (2010). Assessment of Antimicrobial Usage and Residues in Commercial Chicken Eggs from Smallholder Poultry Keepers in Morogoro Municipality, Tanzania. *Zoonoses and Public Health*, 57, 339–344. doi:10.1111/j.1863-2378.2008.01226.x.
- Nouri, N. V., & Salehi, A. (2023). Investigation of the antibiotic residues of broiler meat in northern Iran. *Journal of the Science of Food and Agriculture Reports*, 4(1), 33–38. <https://doi.org/10.1002/jsf2.178>.
- Okombe, E. V., Luboya, W. L. R., Nzuzi, M. G., & Pongombo, S. C. (2016). Détection des résidus d'antibiotiques dans les denrées alimentaires d'origine bovine et aviaire commercialisées à Lubumbashi (RD Congo). *Journal of Applied Biosciences*, 102, 9763–9770.

- Orounladji, B. M., Oke, O. F., Tozo, S. K., & Chrysostome, C. A. A. M. (2022). Socioeconomic Correlates, Typology and Characterization of Indigenous Guinea Fowl (*Numida meleagris* Linnaeus) farming in Benin, West Africa. *Heliyon*, 8, 1–10. <https://doi.org/10.1016/j.heliyon.2022.e09226>.
- Oufella, M., & Smail, F. (2012). *Détection des résidus d'antibiotiques dans la viande du poulet de chair*. Mémoire de Fin de Cycle en Vue de l'Obtention du Diplôme d'Ingénieur d'Etat en Contrôle de Qualité et Analyse. Université Abderrahmane MIRA de Bejaia, Faculté des Sciences de la Nature et de la Vie. 69p.
- Oyededeji, A. O., Msagati, T. A. M., Williams, A. B., & Benson, N. U. (2021). Detection and quantification of multiclass antibiotic residues in poultry products using solid-phase extraction and high-performance liquid chromatography with diode array detection. *Heliyon*, 7, 11p. <http://creativecommons.org/licenses/by-nc-nd/4.0/>.
- Pare, N. G. (2012). *Contribution a l'étude de l'utilisation des médicaments vétérinaires dans les élevages avicoles modernes de la zone périurbaine de Dakar (Sénégal)*. Thèse présentée et soutenue pour obtenir le grade de Docteur Vétérinaire (Diplôme d'Etat). Faculté de Médecine, de Pharmacie et d'Odonto-Stomatologie de Dakar, Sénégal. 113 p.
- Parent, E. (2009). *Détermination quantitative d'antibiotiques (chlortétracycline, oxytétracycline, tylosine) dans quelques types de fumiers de ferme enrichis artificiellement*. Mémoire de maîtrise présenté dans le cadre du programme de maîtrise en Microbiologie agricole pour l'obtention du grade de Maître ès sciences (M.Sc.). Département des Sols et e Génie Agroalimentaire, Faculté des Sciences de l'Agriculture et de l'Alimentation, Université Laval, Québec. 147 p.
- Prajapati, M., Ranjit, E., Shrestha, R., Shrestha, S. P., Adhikari, S. K., & Khanal, D.R. (2018). Status of Antibiotic Residues in Poultry Meat of Nepal. *Nepalese Veterinary Journal*, 35, 55–62.
- Rahmatallah, N., El Rhaffouli, H., Amine, I. L., Sekhsokh, Y., Fihri, O. F., & El Houadfi, M. (2018). Consumption of antibacterial molecules in broiler production in Morocco. *Veterinary Medicine and Science*, 4, 80–90. DOI:10.1002/vms3.89.
- Ramatla, T., Ngoma, L., Adetunji, M., & Mwanza, M. (2017). Evaluation of Antibiotic Residues in Raw Meat Using Different Analytical Methods. *Antibiotics*, 6(34). DOI:10.3390/antibiotics6040034.
- Renaud, D. (2012). Mise au point d'une méthode de dépistage des antibiotiques dans le miel par chromatographie liquide couplée à la spectrométrie de masse en tandem. Chimie analytique, Université de Nantes. <https://hal-anses.archives-ouvertes.fr/anses-00750916>.
- Sangeda, R. Z., Baha, A., Erick, A., Mkumbwa, S., Bitegeko, A., Sillo, H. B., Fimbo, A. M., Chambuso, M., & Mbugi, E. V. (2021). Consumption Trends of Antibiotic for Veterinary Use in Tanzania: A Longitudinal Retrospective Survey From 2010-2017. *Frontiers in Tropical Diseases*, 2:694082. 10p. doi:10.3389/fitd.2021.694082.
- Sani, A. A., Rafiq, K., Hossain, M. T., Akter, F., Haque, A., Hasan, M. I., Sachi, S., Mustari, A., Islam, M. Z., & Alam, M. M. (2023). Screening and quantification of antibiotic residues in poultry products and feed in selected areas of Bangladesh. *Veterinary World*, 16(8), 1747–1754. www.doi.org/10.14202/vetworld.2023.1747-1754.
- Swai, E. S., Sanka, P. N., & Kaaya, J. E. (2013). An investigation of the common causes of death in chicken around Arusha Municipality area, Tanzania. *Livestock Research for Rural Development*, 25(11). <http://www.lrrd.org/lrrd25/11/swai25204.html>.
- Taylor, J. H., & Walker, E. A. (1956). Antibiotics in the preservation of poultry. Paper given at a conference of the United Kingdom Branch of the World's Poultry Science Association, held in London, England, December 6, 1956. pp. 11-18.
- Tek, Z. S., Demirhan, B., & Er Demirhan, B. (2024). Investigation of Quinolone Residues and Total Aerobic Mesophilic Bacteria in Some Poultry Meat and Chicken Eggs. *Bulletin of University of Agricultural Sciences and Veterinary Medicine Cluj-Napoca. Food Science and Technology*, 81(1), 121–129. DOI:10.15835/buasvmcn-fst:2023.0041.
- Ulomi, W. J., Mgaya, F. X., Kimera, Z., & Matee, M. I. (2022). Determination of Sulphonamides and Tetracycline Residues in Liver Tissues of Broiler Chicken Sold in Kinondoni and Ilala Municipalities, Dar es Salaam, Tanzania. *Antibiotics*, 11(1222), 11p. <https://doi.org/10.3390/antibiotics11091222>.
- Vana, T. T. H., Yidanaa, Z., Smookera, P. M., & Coloec, P. J. (2020). Antibiotic use in food animals worldwide, with a focus on Africa: Pluses and minuses. *Journal of Global Antimicrobial Resistance*, 20, 170–177. <http://dx.doi.org/10.1016/j.jgar.2019.07.031>.

- Wouembe, F. D. K. (2013). *Analyse de l'usage des antibiotiques dans les élevages avicoles modernes de poules pondeuses de la région de l'Ouest du Cameroun*. Thèse présentée et soutenue publiquement pour obtenir le grade de Docteur en Médecine Vétérinaire. Faculté de Médecine, de Pharmacie et d'Odontologie de Dakar, Université Cheikh Anta Diop de Dakar (Sénégal). 126p.
- Zamouma, R., Cheblic, A., Djerroud, D., & Alamira, B. (2020). Analyse des quinolones et des sulfamides dans la viande de poulet par CL-SM/SM. *Toxicologie Analytique & Clinique*, 33, 64–73. <https://doi.org/10.1016/j.toxac.2020.09.076>.
- Zerbo, L. H. (2014). *Etude préliminaire sur l'utilisation des antibiotiques dans les élevages de poules pondeuses et la présence de résidus d'antibiotiques dans les œufs commercialisés à Ouagadougou (Burkina Faso)*. Mémoire de Master Qualité des aliments de l'homme, Spécialité : Produits d'origine animale. Présenté et soutenu à l'École Inter-États des Sciences et Médecine vétérinaires (EISMV) de Dakar, Sénégal. 47 p.